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# An outbreak of feline infectious peritonitis in a Taiwanese shelter: epidemiologic and molecular evidence for horizontal transmission of a novel type II feline coronavirus

Ying-Ting Wang<sup>1</sup>, Bi-Ling Su<sup>2</sup>, Li-En Hsieh<sup>1</sup> and Ling-Ling Chueh<sup>1\*</sup>

## Abstract

Feline infectious peritonitis (FIP) is a fatal disease caused by feline coronavirus (FCoV) infection. FCoV can be divided into serotypes I and II. The virus that causes FIP (FIPV) is believed to occur sporadically and spread infrequently from cat to cat. Recently, an FIP outbreak from an animal shelter was confirmed in Taiwan. FCoV from all the cats in this shelter were analyzed to determine the epidemiology of this outbreak. Thirteen of 46 (28.2%) cats with typical signs of FIP were identified. Among them, seven cats were confirmed by necropsy and/or histopathological examinations. Despite the fact that more than one FCoV was identified in this multi-cat environment, the eight FIP cats were invariably found to be infected with a type II FCoV. Sequence analysis revealed that the type II FIPV detected from fecal samples, body effusions and granulomatous tissue homogenates from the cats that succumbed to FIP all harbored an identical recombination site in their *S* gene. Two of the cats that succumbed to FIP were found to harbor an identical nonsense mutation in the *3c* gene. Fecal shedding of this type II virus in the effusive form of FIP can be detected up to six days before death. Taken together, our data demonstrate that horizontal transmission of FIPV is possible and that FIP cats can pose a potential risk to other cats living in the same environment.

## Introduction

Feline infectious peritonitis (FIP) is a fatal disease in cats caused by feline coronavirus (FCoV). FCoV is an enveloped, positive-stranded RNA virus belonging to genus *Alphacoronavirus*, family *Coronaviridae*, within the order *Nidovirales*. The genome size of FCoV is approximately 28.9 kb, including a non-structural replicase gene; four structural genes encoding the spike (S), envelope, membrane and nucleocapsid proteins; and five accessory genes *3abc* and *7ab* [1].

Feline coronaviruses cause mild to inapparent and transient infections of the gut and are ubiquitous in cat populations worldwide [2]. They exist in two serotypes, I and II [3]. Type I FCoV is predominant in the field, whereas type II virus represents only 2-30% of infection

[4-8]. Accumulating genetic evidence indicates that type II FCoV have arisen by two homologous recombinations between type I FCoV and canine CoV (CCoV) [9,10]. Both serotypes can mutate in the host to acquire macrophage tropism and cause a systemic disease known as feline infectious peritonitis [2,11,12]. Due to a lack of virus-shedding in studies of FIP cats, the mutant FIP viruses (FIP causing FCoV, FIPV) are presumably contained only within the diseased tissues and not transmitted by cat-to-cat contact under natural circumstances [2,11,13,14].

In this paper, we report herein on an epizootic of FIP in a Taiwanese shelter that was caused by a novel type II FCoV. Epidemiological and molecular studies of isolates from various healthy and affected cats in this shelter strongly suggest that this virus was brought in by the introduction of kittens from another shelter with subsequent horizontal spread to co-housed adult cats.

\* Correspondence: [linglingchueh@ntu.edu.tw](mailto:linglingchueh@ntu.edu.tw)

<sup>1</sup>Graduate Institute of Veterinary Medicine, School of Veterinary Medicine, National Taiwan University, Taipei 10617, Taiwan

Full list of author information is available at the end of the article

## Materials and methods

### Animals and specimen collection

A total of 46 cats from a private cat shelter were subjected to this study from September 2011 to August 2012. The shelter houses adult cats and occasionally some kittens. All of the cats were either stray animals or had been rescued, and some of them were being temporarily raised at the homes of various private cat rescuers. Before this outbreak, all cats were living together in a cageless indoor environment and shared food, water bowls and litter boxes. Some of the cats were siblings, and the others were unrelated (Table 1).

Feces or rectal swab samples of all asymptomatic cats were collected at least once to monitor the presence of FCoV. For cats showing signs of illness suspected to be FIP, body effusions, whole blood and swab samples, including rectal, nasal, oral and conjunctival swabs, were collected routinely. In addition to supportive care, these FIP-suspected animals were subjected to treatment with prednisolone (PreLon<sup>®</sup>, YF Chemical Corp., New Taipei City, Taiwan), benazepril (Cibacen<sup>®</sup>, Novartis, Barbera del Valles, Spain) and recombinant human interferon alpha (Roferon<sup>®</sup>-A, Roche, Basel, Switzerland). Cats that succumbed to illness were subjected to necropsy for pathological confirmation. Upon necropsy, body effusions were first drawn by needle and syringe, followed by collecting swabs, whole blood, urine and granulomatous lesions in the internal organs. All samples were frozen at -20 °C until use. All specimens were screened for FCoV by reverse transcription-nested polymerase chain reaction (RT-nPCR) [15]. The specimens that tested positively were subsequently subjected to further analysis.

### Sample preparation and reverse transcription

Swab samples were suspended in 1 mL of 0.1% diethyl pyrocarbonate (DEPC)-treated water. Fecal samples were suspended with 9 times of 0.1% DEPC-treated water by vortexing. The suspension was centrifuged, and the supernatant was transferred into a new tube. Approximately 0.5 g tissues were frozen and then ground with a mortar and pestle in the presence of 2 mL of Trizol [16]. Total RNA was extracted from 300 µL of swab suspension, whole blood, fecal suspension, tissue homogenate and body effusion using Trizol. Twenty-one microliters of isolated RNA were reverse transcribed with template-specific primer N1 (5'-gctacaattgtatcctcaac-3') or P211 [15] with Moloney murine leukemia virus reverse transcriptase (Invitrogen, CA, USA). The reaction was incubated at 37°C for 60 min, 72°C for 15 min and finally, at 94°C for 5 min.

### Typing of FCoV using nested PCR

For typing of FCoV, nested PCR was performed according to the procedures reported by Addie et al. [5]

with slight modification. Following reverse transcription, 5 µL of the complementary DNA was added to 25 µL of the PCR mixture (Invitrogen, CA, USA) according to the manufacturer's instructions for the following primer sets: S1 and Iffs for type I FCoV detection and S1 and Icfs for type II FCoV detection. Nested PCR was performed on 2 µL of the first PCR product using nested primers. The expected size of the second PCR yielded for type I and II FCoV were 360 and 218 bp, respectively. The products of RT-nPCR were electrophoresed, and then, target DNA fragments were purified (Geneaid Biotech, Ltd, Taipei) and sequenced (Mission Biotech, Taipei, Taiwan) from both orientations.

### Amplification, sequencing and analysis of 3a and 3c gene from the type II FCoV

To amplify the 3a gene of type II FCoV from FIP cats, a specific primer set that can amplify from the type II S gene to the 3a gene was designed. Complementary DNA amplified with the primer set targeted the 3' end of the type II FCoV S gene (Icfs) and 5' end of the FCoV 3a gene (3aR2: 5'-caccaaacctatacacacaag-3'). The temperature cycling consisted of 5 min of preheating at 94°C; 35 cycles of denaturation at 94°C for 20 s, annealing at 50°C for 20 s and extension at 72°C for 30 s; and a final extension at 72°C for 5 min. Following a second round of amplification with the primers nIcfs and 3aR2, the expected size of the product was approximately 600 bp. The amplicons were electrophoresed, purified and sequenced from both orientations to confirm the nucleotide sequences.

To amplify the 3c gene of type II FCoV from FIP cats, specific primer sets that can amplify from the 3' end of the type II S gene to the 3c gene were designed. Complementary DNA were amplified with the forward primer (Icfs) and reverse primer (E68R: 5'-aatatcaatataattatctgctgga-3' and/or N21R: 5'-gttcatctccccagttgacg-3'). The temperature cycling consisted of 5 min of preheating at 94°C; 40 cycles of denaturation at 94°C for 30 s, annealing at 46°C for 30 s and extension at 72°C for 90 s; and a final extension at 72°C for 7 min. Following a second round of amplification with primer nIcfs and E68R, the products were electrophoresed, purified and sequenced from both orientations to confirm the nucleotide sequences.

### Phylogenetic analysis and recombination site analysis of a type II FCoV

Multiple sequence alignments were preformed with ClustalW 2.0 and manually edited in EditSeq (DNASTAR, Madison, USA). Phylogenetic analyses were conducted using MegAlign, version 7.2.1 (DNASTAR, Madison, USA). Bootscan and similarity graphs were prepared with SimPlot 3.5.1 software (SCRsoftware, Baltimore, USA).

**Table 1 Information for all FIP-suspected and confirmed cats from the cat shelter**

Cat no	Age <sup>1</sup>	Date of enter the shelter	Date of fever onset	Date of death	Clinical findings	Necropsy findings	Effusive/non-effusive
1	3m	Jun. 16, 2011	Aug. 17, 2011	Sep. 01, 2011	Fever, anorexia, ascites, neurological signs		
2 <sup>a</sup>	4m	Aug. 06, 2011	NA <sup>2</sup>	Sep. 21, 2011	Clinical signs unavailable		
3 <sup>b</sup>	3m	Jul. 11, 2011	Aug. 18, 2011	Sep. 25, 2011	Fever, anorexia, weight loss, neurological signs		
4	2.5m	Jun. 08, 2011	Aug. 16, 2011	Sep. 28, 2011	Fever, ascites, neurological signs		
5 <sup>a</sup>	4m	Aug. 06, 2011	Aug. 15, 2011	Oct. 20, 2011	Fever, pleural effusion, diarrhea		
6	7m	Apr. 24, 2011	NA	Oct. 22, 2011	Anorexia, weight loss, neurological signs		
7	3y6m	Resident	NA	Oct. 27, 2011		Ascites, jaundice, granulomatous lesion in kidney, fibrinous peritonitis	Effusive
8	6m	Jul. 11, 2011	NA	Dec. 14, 2011		Granulomatous changes in kidney, liver, lung, brain and eyes	Non-effusive
9	2y	Resident	NA	Dec. 28, 2011		Ascites, pleural effusion and pericardial effusion, granulomatous changes in kidney, liver and intestine	Effusive/non-effusive
10 <sup>b</sup>	3m	Jul. 11, 2011	NA	Nov. 05, 2011		Granulomatous changes in kidney, liver and omentum	Non-effusive
11 <sup>c</sup>	1y6m	Resident	NA	Feb. 14, 2012		Ascites and pleural effusion, jaundice, fibrinous peritonitis, granulomatous changes in kidney, liver, lung and spleen.	Effusive/non-effusive
12 <sup>c</sup>	1y6m	Resident	NA	Mar. 19, 2012		Jaundice, fibrinous peritonitis, granulomatous changes in thoracic and abdominal wall, kidney, liver, lung, spleen omentum, and eyes.	Effusive/non-effusive
13	1y7m	Resident	NA	Apr. 13, 2012		Jaundice, enlargement of liver and mesenteric lymph node, granulomatous changes in kidney and lung.	Non-effusive

<sup>1</sup> Age of the cats when clinical signs of FIP appeared.

<sup>2</sup> Not available.

<sup>a, b, c</sup>: Siblings.

## Results

### Confirmation of an FIP outbreak in the cat shelter

The shelter had been ongoing for three-and-a-half years. No history of FIP was recorded before August 2011. Kittens (cat 1, 3, 4, 8 and 10) were introduced to this shelter between June and July 2011. After entering, these kittens played together and lived with the resident adult cats. Before the outbreak, the kittens were taken individually to a veterinarian for vaccinations and attended adoption activities. Fever was first observed in four of the kittens (cat 1, 3, 4, 5) within a period of a few days (from August 15<sup>th</sup> to 18<sup>th</sup>) (Table 1). Clinical signs, e.g., fever, anorexia, neurological signs, panting and/or abdominal extension were observed for the following two months and the kittens died sequentially between September 1<sup>st</sup> and October 22<sup>nd</sup> (Table 1). The caregivers at the shelter requested our help on September 27<sup>th</sup>. All of the resident cats in the shelter were immediately screened for FCoV using RT-nPCR. All FCoV positive cats were isolated individually and kept separated. Nevertheless, starting in September, some of the adult cats with FIP (cats 7-13) showed clinical signs similar to those of the kittens and all of them died later.

Six kittens (cats 1-6) with body effusions and/or neurological signs succumbed within the first two months without necropsy confirmation (Table 1). Cat 1 was once presented to our teaching hospital, and ascites was taken from the cat. In cats 7-13, typical necropsy features, characterized by ascites and/or pleural effusions in the body cavities (effusive FIP) and granulomatous lesions in several organs, especially the kidney, liver, lung, omentum and eyes (non-effusive FIP), were found. Cats 9, 11 and 12 manifested mixed forms of the disease at necropsy (Table 1).

Altogether, 13 of 46 cats (28.3%) died from FIP between September 2011 and April 2012. During this period, 33 cats (71.7%) appeared to be clinically healthy, and 26 of these asymptomatic cats (78.7%) were positive at least once for FCoV detection in their feces by RT-nPCR. The other seven of these asymptomatic cats were negative for FCoV detection (Table 2).

### Type II FIPV were consistently detected in the FIP-succumbed cats

To further investigate the relationship between these seven histopathologically confirmed FIP cats, amplified DNA were typed, sequenced and analyzed. Type II FIPV were detected in all eight of the animals that succumbed to FIP in their swab samples, feces, urine, body effusions, cerebrospinal fluids and tissue homogenates (Table 3). Type II viruses that cause FIP were found not only in the diseased tissue but also in the fecal samples (cat 7, 11, 12 and 13), nasal/oral/conjunctival swab samples (cat 7, 8, 9, 11 and 12), and urine collected by cystocentesis

**Table 2 Detection and typing of FCoV in fecal samples from healthy cats in the same shelter**

No.	FCoV				Type
	Oct. 2011	Feb. 2012	Jun. 2012	Jul. 2012	
14	++	++	++	+	untypable
15	-	+	-		untypable
16	++	-	-		untypable
17	++	++	+	++	I
18	++	++	++	++	I
19	-		-	+	untypable
20	-		-	+	untypable
21	-		-	-	
22	++	++	-		untypable
23	+	++	-	+	I
24	-	+	-		untypable
25	++	++	+	+	I
26	-		+	-	untypable
27	++	++	+	+	I
28	++	++	++	+	I
29	-		-	-	
30	-	++	++	-	I
31	-			-	
32	+	++	-	-	I
33	-		++	-	untypable
34	++				I
35	-		+	+	untypable
36	++	++	+	-	I
37			-		
38			+		untypable
39		++	+	+	I
40		-	+	-	untypable
41		+	-	+	untypable
42			+	-	untypable
43			-		
44				-	
45				-	
46			+		untypable

++: FCoV detected in the first round of PCR.

+: FCoV detected only in the nested PCR.

(cat 11) (Table 3). Although there was no necropsy, the ascites from cat 1, the first FIP death in this cat shelter, was available for analysis. This cat was confirmed to be infected with a type II virus. From the healthy animals, only type I or untypable FCoV were detected from the fecal samples (Table 2). Cats 8, 9 and 13 were co-infected with both types of FCoV (Table 3). Although more than one FCoV, i.e., type I, II or untypable viruses, was found to circulate in this multi-cat environment, the

**Table 3 The characteristics of 3c genes of FCoV recovered from various specimens of FIP cats**

Case no.	Genotype of FCoV											S gene crossover site	Integrity of the 3c gene <sup>b</sup>							
	NOC	R/F	U	A/P	CSF	Li	Lu	Ki	Br	Sp	Int		R/F	A/P	Li	Lu	Ki	Br	Sp	
1												4250 <sup>a</sup>		intact						
7												4250	intact		intact	intact	intact		intact	
8						+			+	-	-									
9						+					+	4250		G210*						
10						+	+					4250						intact		
11					+					+		4250			E47*					
12						+					+	4250	G210*	G210*						
13		/			+	+	+			+		4250						Q218*		

NOC, nasal/oral/conjunctival swabs; R/F, rectal swabs or fecal samples; A/P, ascites or pleural effusions; CSF, cerebrospinal fluid; Li, liver; Lu, lung; Ki, kidney; Br, brain; Sp, spleen; Int, intestine.

+: FCoV positive but the type of virus cannot be determined.

-: FCoV negative.

<sup>a</sup>: FCoV/NTU2/R/2003; GenBank: DQ160294.

<sup>b</sup>: E47\*, G210\* and Q218\*: truncated 3c proteins with premature stop codons at amino acids 47, 210 and 218 were found, respectively.

eight FIP cats were consistently found to be infected with a type II FCoV, whereas the healthy cats were not (Tables 2 and 3).

#### A type II FIPV of the same origin was detected from FIP-succumbed cats

To further investigate the relationship of these disease-associated type II viruses isolated from FIP-succumbed cats, specific primer sets that can specifically amplify from the 3' end of type II S gene to a downstream gene were used to analyze the viral sequences. The identity of the 620 bp amplicons derived from seven type II FIPV was approximately 98.7% to 99.8%. Phylogenetic analysis revealed that the type II FCoV derived from this outbreak were all grouped in a separate cluster, distinct from the other four type II FCoV currently available in the GenBank, i.e., FIPV 79-1146 (GenBank: DQ010921), FCoV 79-1683 (GenBank: JN634064), FCoV DF-2 (GenBank: DQ286389) and FCoV NTU156 (GenBank: GQ152141) (data not shown).

A recombination event at the 3' end of the S gene with the putative recombination site located at nucleotide 4250 was identified from all of the type II FCoV derived from the body effusions and tissue homogenate of cats 1, 7, 9, 10, 11, 12 and 13 (Additional file 1) (Table 3). Sequences upstream from this site show a higher similarity to CCoV, whereas sequences downstream from this site were more similar to type I FCoV (Figure 1). These findings highly suggest that the type II FCoV recovered from all of the FIP cats were from a common origin.

#### An identical nonsense mutation in 3c gene was found in two FIP-succumbed cats

To further analyze the relationship of these FIPV, 3c genes, the proposed virulence-associated factor of FIP,

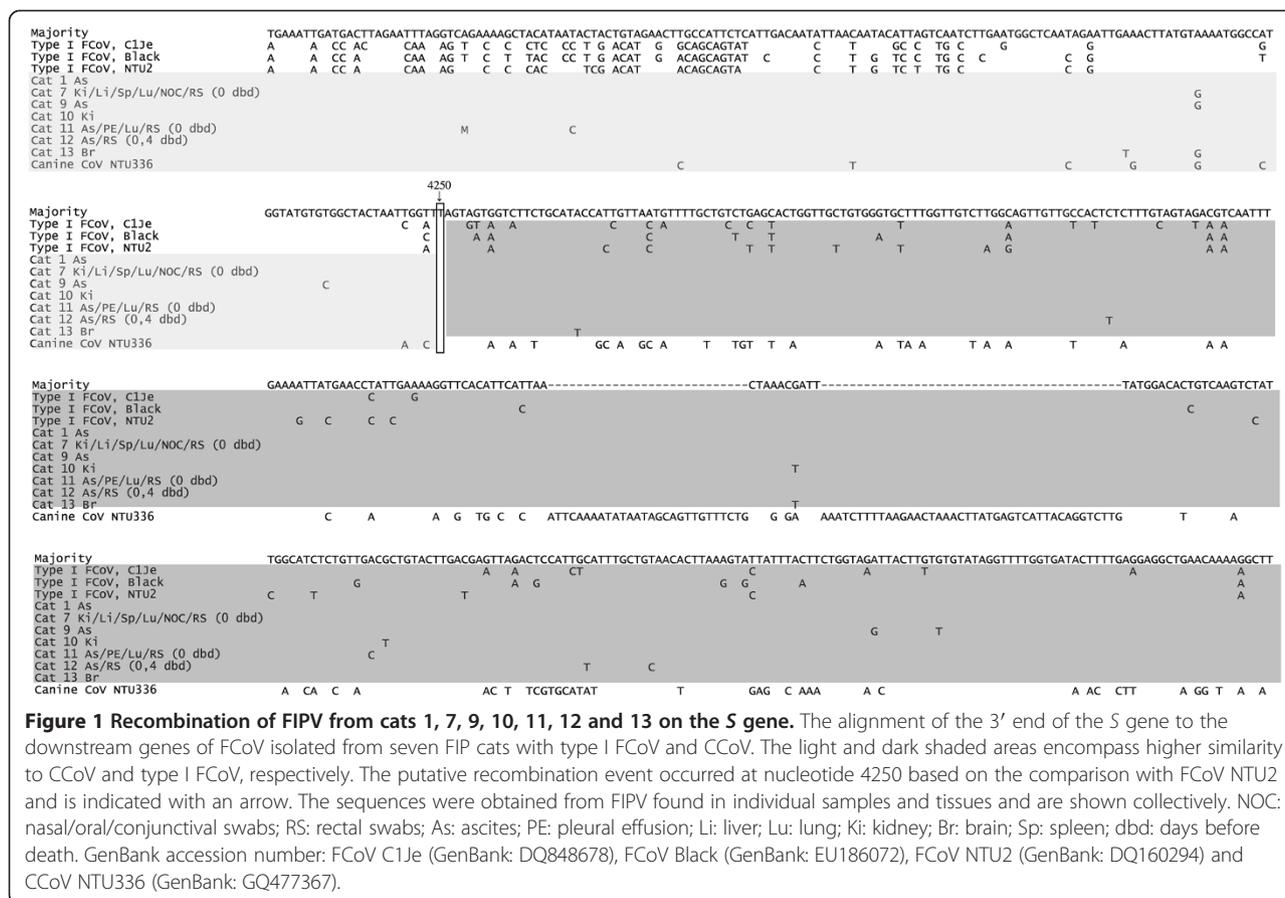
were amplified from disease-related type II FCoV. Mutated 3c genes with an identical premature stop codon at nucleotides 628-630 (amino acid 210, G210\*) were found in two FIP cats, including cats 9 (ascites, spleen and brain) and 12 (ascites and rectal swabs from the day it succumbed and four days before) (Figure 2A). It is noteworthy that the FIPV shed from cat 12 harbored an identical nonsense mutation as the virus in its ascites. Intact 3c genes were detected from cats 1, 7 and 10, which had previously succumbed to FIP. Two more distinct nonsense mutations were identified from cats 11 (E47\*) and 13 (Q218\*) (Figure 2A-B, Table 3).

#### Shedding of the type II FIPV can be detected at the terminal stage of FIP cats

The presence of FCoV was continuously analyzed to elucidate the possible route of shedding and transmission of FIPV. Disease-associated type II FCoV were found to shed through nasal/oral/conjunctival and fecal routes (Table 4). Fecal and nasal/oral/conjunctival shedding of this type II virus can be detected from the 6<sup>th</sup> day (cat 11) and 4<sup>th</sup> day (cat 12) respectively, before death. Viremia could be detected during the terminal stage in the cats with FIP, up to 18 days before death, and simultaneous fecal shedding was noted in one cat (cat 12) (Table 4).

#### Discussion

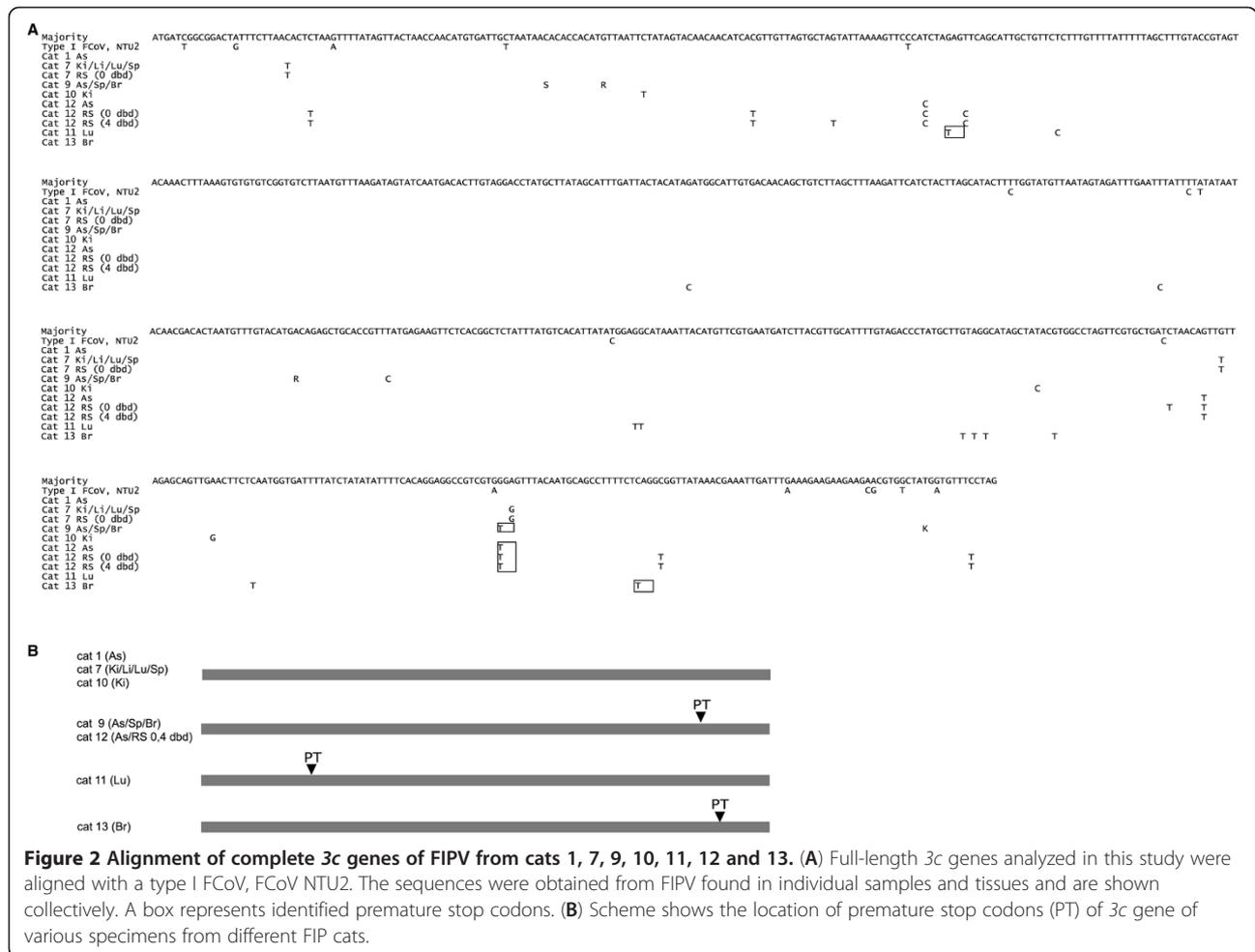
The possibility of horizontal transmission in FIP is generally questioned because (i) the occurrence of FIP is sporadic, and a single cat developing FIP in a multi-cat environment is common [2]; (ii) the internal mutation theory, which describes that FIPV is a mutant generated from enteric FCoV in one cat [12,17]; (iii) there is a lack of evidence of mutant FIPV shedding from FIP cats; and



(iv) the mutation of the 3c gene is unique to each FIP cat [11,13,18]. The current belief is that cats that succumb to FIP do not shed and transmit the FIPV to other cats [11,13,14,18-20]. Our data indicate that this outbreak of FIP was caused by viruses from the same origin. First, all of the FIP losses involved a type II FIPV infection, and the recombination event of these seven type II viruses was mapped to the same site. The recombination sites of the type II viruses currently available in GenBank, i.e., FIPV 79-1146, FCoV 79-1683 and FCoV NTU156, were all unique and occurred independently [9,10]. Second, the FIPV identified in the three kittens that died within the first two months after the onset of fever harbored an intact 3c gene, whereas the viruses from the cats that survived longer (died four to eight months later) all bore a nonsense mutation, i.e., G210\* (cat 9 and 12), E47\* (cats 11) and Q218\* (cat 13). Because the three nonsense mutations detected in the FIPV from these animals were all located at different sites, the viruses that originally infected these cats should all harbor an intact 3c gene - like the virus recovered from the kittens that succumbed earlier. After infection, point mutations arose during the replication of the virus in the individual cats, giving rise to FIPV with a 3c gene

bearing nonsense mutations at different locations. The finding that the viruses that were identified not only in the tissues but also in the fecal samples of two cats (cat 9 and 12) harbored an identical mutation in the 3c gene further confirmed that horizontal transmission had occurred (Table 3). Taken together, all these findings demonstrated that a highly virulent FIPV had spread from animal to animal in a horizontal manner.

This is the first report of a type II FIPV outbreak with evidence of horizontal transmission of a disease-associated FCoV. The outbreak of FIP started after introducing five kittens (cat 1, 3, 4, 8 and 10) into this shelter between June and July of 2011. Because the causative type II viruses with the distinct genetic marker in the S gene were confirmed to be a recombinant of feline and canine coronaviruses and some of the kittens that died earlier were found to have been living closely with dogs between being rescued and entering the shelter, any of these kittens could have been the source of this type II virus. Dogs, and especially young dogs, in shelters often shed large amounts of canine coronavirus in their feces and recombination between cat and dog and dog and pig coronaviruses has been well documented [21-23]. Additionally, these causative type II viruses



**Figure 2 Alignment of complete 3c genes of FIPV from cats 1, 7, 9, 10, 11, 12 and 13. (A)** Full-length 3c genes analyzed in this study were aligned with a type I FCoV, FCoV NTU2. The sequences were obtained from FIPV found in individual samples and tissues and are shown collectively. A box represents identified premature stop codons. **(B)** Scheme shows the location of premature stop codons (PT) of 3c gene of various specimens from different FIP cats.

**Table 4 Shedding and serotypes of feline coronavirus detected in FIP cats in the cat shelter**

Cat no.	Sample	Days before death																	
		-80	-66	-60	-57	-50	-43	-36	-29	-25	-23	-20	-18	-14	-12	-8	-6	-4	0*
9	Feces																		
	NOC swabs																		
	Viremia																		+
	Effusions																		
11	Feces													-					
	NOC swabs													-				-	
	Viremia													-	-		-	-	-
	Effusions													+					
12	Feces	-	+	-												+			
	NOC swabs																		
	Viremia	-		-											+	+		-	
	Effusions																		

+: FCoV positive; -: FCoV negative.

I, II: Type I or type II FCoV.

\*: Samples were collected directly prior to euthanasia, except for cat 12, from which the samples were taken after death.

were detected in many excretions and secretions of cats dying of FIP (Table 3), thus providing a vehicle for cat-to-cat spreading.

Although directly following the first FCoV screening of all the animals in this shelter, cats shedding FCoV were caged separately, and transmission subsequently ceased, the mortality rate for this outbreak was high (28%, 13/46). Three studies regarding outbreak of FIP have been reported previously. In a four-year study conducted in a closed breeding cattery, the average mortality was 17.3% [24], and in a 10-year study of a closed breeding colony, the mortality was 29.4% (5/17) [25]. Another epidemic study conducted in seven catteries/shelters revealed a > 10% mortality rate [20]. The high incidence of FIP in those closed breeding colonies might be influenced by the genetically predisposed breeding stock. In our study, only several FIP cats in this shelter were siblings, and the others were genetically unrelated. Our study demonstrates that without the influence of genetic predisposing factors, the mortality of FIP can still be high in a closed multi-cat environment as long as the spread of the disease-associated FCoV remains undetected.

In this multi-cat environment, three of the FIP cats were infected with not only type II FCoV but also co-infected with type I FCoV (Table 3). Type I FCoV were found only in the fecal samples, whereas type II FCoV were found in the diseased-associated samples, including body effusion, granulomatous tissue homogenates and cerebrospinal fluid. This finding indicates that in these dually infected animals, type II FCoV was the main cause of FIP. This finding is consistent with our previous finding that infection with type II FCoV appears to be significantly correlated with FIP [4].

The presence of FCoV in the whole blood at the terminal stage has been observed previously [26,27]; however, to our knowledge, fecal shedding of FIPV before the end stage of the disease has not been reported prior to this study. Shedding of this type II virus through fecal and nasal/oral/conjunctival routes can be detected in effusive form FIP up to six days before death. Another experimental infection study showed that inoculated viruses could only be recovered for approximately two weeks after inoculation, before clinical signs of disease develop [14]. Taken together, the transmission of FIPV might occur at the onset, before disease manifestation and at the terminal stage. In this outbreak, all the cats were housed together in an open room at the very beginning. After seven cats succumbed sequentially, all FCoV-positive cats were caged individually, and kept separated. The isolation eventually terminated the transmission of the disease. This outbreak, which killed 13 cats, allowed us to determine clearly that FIPV could be transmitted horizontally and showed that isolation of

diseased cats should be taken into consideration in a multi-cat environment.

## Additional file

**Additional file 1: Recombination site analysis of FIPV from cats 1, 7, 9, 10, 11, 12 and 13 on the S gene.** Similarity plot analysis with the Kimura (two-parameter) distance model, neighbor-joining tree model and 100 bootstrap replicates showed a recombination event, and the putative crossover site is indicated with an arrow.

## Competing interests

The authors declare that they have no competing interests.

## Authors' contributions

YTW performed the sampling and preparation, FCoV detection, typing, 3c gene amplification and further analysis and prepared the manuscript. BLS supervised the sampling and treatment of all FIP animals and contributed to the preparation of the manuscript. LEH participated in 3c gene amplification, genetic analysis and the preparation of the manuscript. LLC conceived the study, participated in study design and coordination and contributed to the preparation of the manuscript. All authors read and approved the final manuscript.

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## Author details

<sup>1</sup>Graduate Institute of Veterinary Medicine, School of Veterinary Medicine, National Taiwan University, Taipei 10617, Taiwan. <sup>2</sup>Graduate Institute of Veterinary Clinical Sciences, School of Veterinary Medicine, National Taiwan University, Taipei 10617, Taiwan.

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